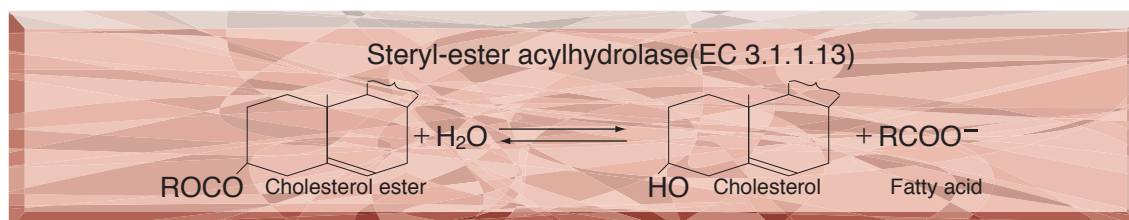


● TOYOBO ENZYMES ●  
(Diagnostic Reagent Grade)

# CHOLESTEROL ESTERASE

*from Microorganism*



## PREPARATION and SPECIFICATION

Appearance	: Light brown amorphous powder, lyophilized
Activity	: Grade III 5.0U/mg-solid or more
Contaminant	: Catalase $\leq 1.0 \times 10^{-2}\%$
Stabilizers	: Mg <sup>++</sup> , BSA

## PROPERTIES

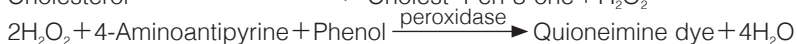
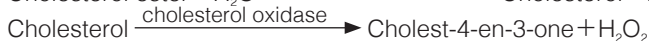
Stability	: Stable at $-20^{\circ}\text{C}$ for at least one year	(Fig.1)
Michaelis constants	: $5.7 \times 10^{-5}\text{M}$ (Linoleate), $7.2 \times 10^{-5}\text{M}$ (Oleate)	
Inhibitors	: Hg <sup>++</sup> , Cu <sup>++</sup>	
Optimum pH	: 6.0	(Fig.2)
Optimum temperature	: $40^{\circ}\text{C}$	(Fig.3)
pH Stability	: pH 5.5–10.0 (25 $^{\circ}\text{C}$ , 20hr)	(Fig.4)
Thermal stability	: below $40^{\circ}\text{C}$ (pH 7.0, 15min)	(Fig.5)
Effect of various chemicals	: (Table 1)	

## APPLICATIONS

This enzyme is useful for enzymatic determination of total cholesterol when coupled with cholesterol oxidase (COO-311, COO-321, COO-331) in clinical analysis.

## ASSAY

### Principle:



The appearance of quinoneimine dye formed by coupling with 4-aminoantipyrine and phenol is measured at 500nm by spectrophotometry.

### Unit definition:

One unit causes the formation of one micromole of hydrogen peroxide (half a micromole of quinoneimine dye) per minute under the conditions described below.

### Method:

#### Reagents

- A. 0.2M K-Phosphate buffer, pH 7.0
- B. Cholesterol linoleate solution : To 39mg of cholesterol linoleate, add 2.0ml of isopropanol and dissolve completely with heating slightly. Mix with about 80ml of 1.0% (v/v) hot Triton X-100 solution (preheated at 72–74°C) to the cholesterol linoleate solution and keep the solution in a hot water bath (72–74°C), with stirring for 30 minutes. The solution will turn clear and then cloudy. Cool under running water with gentle agitation until temperature of the solution goes down to room temperature. Add 600mg of Na-cholate and dissolve. Fill up the solution to 100ml with 1.0% Triton X-100 solution. This solution is stable at 4°C for at least 5 days.
- C. 4-AA solution : 1.76% (1.76g 4-aminoantipyrine/100ml of H<sub>2</sub>O)(Store at 4°C in a brownish bottle)
- D. Phenol solution : 6.0% (6.0g phenol/100ml of H<sub>2</sub>O)(Store at 4°C in a brownish bottle)
- E. POD solution : Horseradish peroxidase (Toyobo, Grade III) 7,500 purpurogallin units/50ml of 0.1M K-phosphate buffer, pH 7.0 (150 PU/ml)(Prepare freshly)
- F. COD solution : Streptomyces sp. cholesterol oxidase (Toyobo, Grade III) 1,500U/5.0ml of ice-cold H<sub>2</sub>O(300 U/ml)(Prepare freshly)
- G. Enzyme diluent : 20mM K-phosphate buffer, pH 7.5 containing 2mM MgCl<sub>2</sub>, 0.5mM EDTA-Na<sub>3</sub> and 0.2% BSA

#### Procedure

- Prepare the following working solution (50 tests) in a brownish bottle.
 

75 ml	Buffer solution	(A)
50 ml	Substrate solution	(B)
2.5ml	4-AA solution	(C)
5.0ml	Phenol solution	(D)
5.0ml	POD solution	(E)

(This solution is stable at 4°C for at least 5 days.)

Concentration in assay mixture	
K-Phosphate buffer	0.11 M
Cholesterol linoleate	0.20mM
4-Aminoantipyrine	1.5 mM
Phenol	22 mM
EDTA	17 μM
Isopropanol	0.68 %
COD	ca.10 U/ml
POD	ca. 5.1 U/ml

- Pipette 2.75ml of working solution into a cuvette (d=1.0cm) and equilibrate at 37°C for about 5 minutes. Add 0.1ml of COD solution (F), mix and keep at 37°C for another 2 minutes.
- Add 0.1ml of the enzyme solution\* and mix with gentle inversion.
- Record the increase in optical density at 500nm against water for 3 to 4 minutes in a spectrophotometer thermostated at 37°C, and calculate the ΔOD per minute from the initial linear portion of the curve (ΔOD test).

At the same time, measure the the blank rate (ΔOD blank) by the same method as test except that the enzyme diluent (G) is added instead of the enzyme solution.

- \* Dissolve the enzyme preparation in ice-cold enzyme diluent (G), and dilute to 0.08–0.22U/ml with the same buffer, immediately before assay.

#### Calculation

Activity can be calculated by using the following formula :

$$\text{Volume activity (U/ml)} = \frac{\Delta \text{OD}/\text{min} (\Delta \text{OD test} - \Delta \text{OD blank}) \times V_t \times d_f}{13.78 \times 1/2 \times 1.0 \times V_s} = \Delta \text{OD}/\text{min} \times 4.282 \times d_f$$

$$\text{Weight activity (U/mg)} = (\text{U/ml}) \times 1/C$$

V<sub>t</sub> : Total volume (2.95ml)

V<sub>s</sub> : Sample volume (0.1ml)

13.78: Millimolar extinction coefficient of quinoneimine dye under the assay conditions (cm<sup>2</sup>/micromole)

1/2 : Factor based on the fact that one mole of H<sub>2</sub>O<sub>2</sub> produces half a mole of quinoneimine dye.

1.0 : Light path length (cm)

d<sub>f</sub> : Dilution factor

C : Enzyme concentration in dissolution (c mg/ml)

## REFERENCES

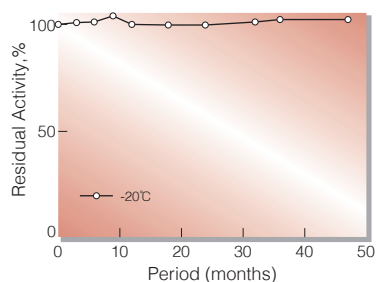
- 1) C.C.Allain, L.S.Poon, C.S.G.Chan, W.Richmond, and P.C.Fu; *Clin.Chem.*, **20**, 470 (1974).
- 2) Y.Kameno, N.Nakano, and S.Baba; *Jap.J.Clin.Path.*, **24**, 650 (1976).

**Table 1. Effect of Various Chemicals on Cholesterol esterase**

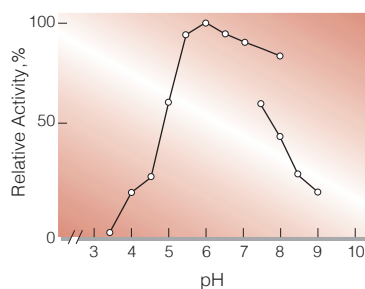
[The enzyme (40U/ml) was incubated at 25°C for 1hr with each chemical.]

Chemical	Concn.(mM)	Residual activity(%)	Chemical	Concn.(mM)	Residual activity(%)
None	—	100	MIA	2.0	95
CaCl <sub>2</sub>	2.0	103	NaF	20.0	101
Ba(OAc) <sub>2</sub>	2.0	93	NaN <sub>3</sub>	20.0	100
FeCl <sub>3</sub>	2.0	97	EDTA	5.0	88
CoCl <sub>2</sub>	2.0	98	o-Phenanthroline	2.0	100
MnCl <sub>2</sub>	2.0	82	$\alpha, \alpha'$ -Dipyridyl	2.0	96
Zn(OAc) <sub>2</sub>	2.0	100	Borate	20.0	99
NiCl <sub>2</sub>	2.0	99	Triton X-100	1.0%	84
Pb(OAc) <sub>2</sub>	2.0	76	Brij 35	1.0%	99
AgNO <sub>3</sub>	2.0	94	SDS	0.1%	91
HgCl <sub>2</sub>	2.0	58	Tween 20	0.1%	98
CdCl <sub>2</sub>	1.0	101	Span 20	0.1%	101
CuSO <sub>4</sub>	1.0	1.5	Na-cholate	1.0%	99
NEM	2.0	101	Taurocholate	0.1%	100
PCMB	2.0	91			

Ac, CH<sub>3</sub>CO; PCMB, p-Chloromercuribenzoate; MIA, Monoiodoacetate; EDTA, Ethylenediaminetetraacetate; NEM, N-Ethylmaleimide; SDS, Sodium dodecyl sulfate; DAC, Dimethylbenzylalkylammonium chloride.

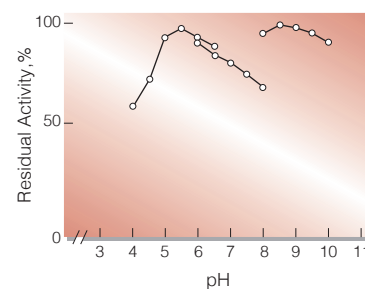


**Fig. 1. Stability (Powder form)**  
[kept under dry conditions]



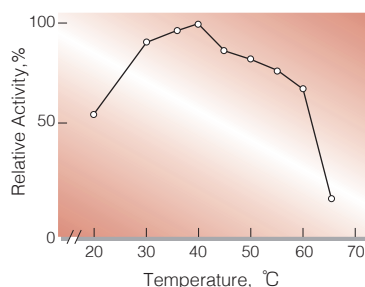
**Fig. 2. pH-Activity**

[37°C in 0.1M buffer solution:  
pH3.5-5.5, acetate; pH5.5-8.0,  
K-phosphate; pH8.0-9.0, Tris-HCl]



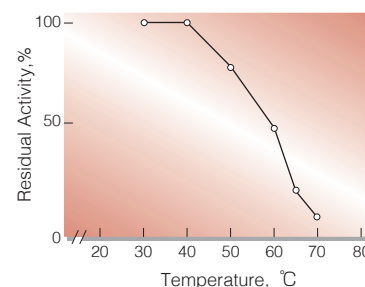
**Fig. 4. pH-Stability**

[25°C, 20hr-treatment with 0.1M  
buffer solution: pH4.0-6.5, acetate;  
pH6.0-8.0, K-phosphate; pH8.0-10.0  
Tris-HCl]



**Fig. 3. Temperature activity**

[in 0.1M K-phosphate buffer, pH7.0]



**Fig. 5. Thermal stability**

[15min-treatment with 0.1M K-phosphate, pH7.5]  
[enzyme concn. 20 U/ml]

## 活性測定法 (Japanese)

### 1.原理

$\text{Cholesterol ester} + \text{H}_2\text{O} \xrightarrow{\text{cholesterol esterase}} \text{Cholesterol} + \text{Fatty acid}$   
 $\text{Cholesterol} \xrightarrow{\text{cholesterol oxidase}} \text{Cholest-4-en-3-one} + \text{H}_2\text{O}_2$   
 $2\text{H}_2\text{O}_2 + 4\text{-Aminoantipyrine} + \text{Phenol} \xrightarrow{\text{peroxidase}} \text{Quinoneimine dye} + 4\text{H}_2\text{O}$

4-AAとフェノールの酸化縮合生成物であるQuinoneimine色素を500nmで測定し、上記反応で生成したH<sub>2</sub>O<sub>2</sub>量(加水分解されたCholesterol esterの量)を定量する。

### 2.定義

下記条件下で1分間に1マイクロモルのcholesterol esterを加水分解する酵素量を1単位 (U)とする。

### 3.試薬

- 0.2M K-リン酸緩衝液, pH7.0
- コレステロールリノレート溶液 [39mgのコレステロールリノレートを精秤し, 2.0mlのイソプロパノールを加え完全に加温溶解する。それを予め72~74°Cに加温しておいた約80mlの1.0%(V/V)トリトンX-100溶液と混和し, 更に72~74°Cの温湯中で攪拌しながら30分間保ったのち, 流水中で室温まで冷却する。次いで600mgのコール酸ナトリウムを加えて溶解させ, 前記1.0%トリトンX-100溶液で最終液量を100mlとする] (4°C保存で5日間は使用可能)
- 1.76%4-AA水溶液 [1.76gの4-アミノアンチピリンを100mlの蒸留水に溶解し, 褐色瓶中で4°Cに保存する]
- 6.0%フェノール水溶液 [6.0gのフェノールを100mlの蒸留水に溶解し, 褐色瓶中で4°Cに保存する]
- POD溶液 [西洋ワサビ由来ペルオキシダーゼ(東洋紡製, GradeⅢ)7,500プルプロガリン単位を50mlの0.1M K-リン酸緩衝液, pH7.0に溶解する(150PU/ml)] (用時調製)
- COD溶液 [ストレプトミセス属由来のコレステロールオキシダーゼ(東洋紡製, GradeⅢ)1,500単位を5.0mlの氷冷蒸留水に溶解する(300 U/ml)] (用時調製)
- 酵素溶液: 酵素標品を予め氷冷した2mM MgCl<sub>2</sub> 0.5mM EDTA-Na<sub>3</sub>及び0.2% BSAを含む20mM K-リン酸緩衝液, pH7.5で溶解し, 分析直前に同緩衝液で0.08~0.22U/mlに希釈する。

### 4.手順

- 下記反応混液を(50テスト)を褐色瓶中で調製する(4°C保存で5日間は使用可能)。
 

75 ml	K-リン酸緩衝液	(A)
50 ml	基質溶液	(B)
2.5ml	4-AA水溶液	(C)
5.0ml	フェノール水溶液	(D)
5.0ml	POD溶液	(E)
- 反応混液2.75mlを試験管に採り, 37°Cで約5分間予備加温し, 0.1mlのCOD溶液を加えて更に2分間加温する。
- 酵素溶液0.1mlを添加し, ゆるやかに混和後, 水を対照に37°Cに制御された分光光度計で500nmの吸光度変化を3~4分間記録し, その初期直線部分から1分間あたりの吸光度変化を求める(ΔOD test)。

- 盲検はCOD添加液②に酵素溶液の代りに酵素希釈液(2mM MgCl<sub>2</sub>, 0.5mM EDTA-Na<sub>3</sub>及び0.2% BSA)を含む20mM K-リン酸緩衝液, pH7.5を0.1ml加え, 上記同様に操作を行って1分間当たりの吸光度変化を求める(ΔODblank)。

### 5.計算式

$$U/ml = \frac{\Delta OD/min (\Delta OD \text{ test} - \Delta OD \text{ blank}) \times 2.95(ml) \times \text{希釈倍率}}{13.78 \times 1/2 \times 1.0 \times 0.1(ml)}$$

$$= \Delta OD/min \times 4.282 \times \text{希釈倍率}$$

$$U/mg = U/ml \times 1/C$$

13.78 : Quinoneimine色素の上記測定条件下でのミ  
リモル分子吸光係数 (cm<sup>2</sup>/micromole)

1/2 : H<sub>2</sub>O<sub>2</sub>の1分子のから形成するQuinoneimine  
色素は1/2分子である事による係数

1.0 : 光路長(cm)

C : 溶解時の酵素濃度(c mg/ml)